



Research Article

**EVALUATION OF ANTI-INFLAMMATORY ACTIVITY OF *CALOTROPIS GIGANTEAN*,
TYLOPHORA INDICA AND *SARCOSTEMMA SECOMONE***

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ABSTRACT

The main objective of the present investigation is to evaluate the anti-inflammatory property of *Calotropis gigantean*, *Tylophora indica* and *Sarcostemma secomone* in Wister rats using the Carrageenan induced left hind paw edema. In the study it was observed that the aqueous, ethanol and chloroform extracts were significantly inhibit paw edema induced by carrageenan in rats. The ethanolic extract showed a significant anti-inflammatory property when compared with the aqueous, chloroform, standard and untreated control. Among the three tested plant species *S.sarcostemma* showed the maximum inhibition in rats.

KEY WORDS: *Calotropis gigantean*, *Tylophora indica*, *Sarcostemma secomone*, anti-inflammatory, Carrageenan.

INTRODUCTION

Inflammation is the complex biological response of vascular tissues to harmful stimuli, such as pathogens, damaged cells, or irritants. It is a protective attempt by the organism to remove the injurious stimuli as well as initiate the healing process for the tissue. Inflammatory responses occur in three distinct temporal phases, each apparently

mediated by different mechanisms: 1) an acute phase characterized by transient local vasodilation and increased capillary permeability; 2) a delayed, sub-acute phase characterized by infiltration of leukocytes and phagocyte cells; and 3) a chronic proliferative phase, in which tissue degeneration and fibrosis occur^{1,2,3,4}. Inflammation is a defense mechanism, the complex events and mediators involved in

the inflammatory reactions induce, maintain or aggravate many diseases^{5,6,7,8}. There are two types of plant chemicals, primary metabolites such as sugars, proteins, amino acids, chlorophylls etc. The other category of chemicals is called secondary metabolites, which includes alkaloids, terpenoids, saponins and phenolic compounds. These chemicals exert a significant physiological effect on the mammalian system. A lot of references are available in the field of ethnomedicinal plants used as anti-inflammatory drugs⁹ have reported the anti-inflammatory activity of two ayurvedic formulations containing 'guggul'. Bhattacharya *et al* have reported anti-inflammatory potential of methanol extract of *Stepenia glabra* of Menispermaceae family.¹⁰ The extract depicted anti-inflammatory activity at the dose of 150 mg/kg body weight. It have revealed the anti-inflammatory activity of bioactive fractions isolated from seeds of *Trigonella foenum gracium* L., roots of *Glycyrrhiza glabra* L. and fruits of *Coriandrum sativum* L.¹¹

However, studies have been continuing on inflammatory diseases indicated that the side effects of the currently available anti-inflammatory drugs pose a major problem during their clinical use^{12,13,14}. Low cost and easy availability are the factors that generated a

renewed interest in plant medicine in the last decade. The traditional practitioners in India prescribe the leaves to the patients without regard to any possible adverse effects in the view of its many uses. Therefore, development of newer, more powerful anti-inflammatory drugs lesser side effects are necessary. Hence the present study planned to evaluate the effect of aqueous, methanol and chloroform extracts of the leaves of *Calotropis gigantea* *Tylophora indica* and *Sarcostemma secomone* on anti-inflammation in rats.

EXPERIMENTAL METHODS

For the evaluation, the leaves of *Calotropis gigantea* *Tylophora indica* and *Sarcostemma secomone* were collected from the area of different parts of Guntur District Andhra Pradesh India. The Plant were identified and authenticated. After cleaning the leaves were shade dried at room temperature for 10 days and coarsely powdered with the help of a hand-grinding mill and the power was sieved. The air-dried and powdered leaves were extracted successively with aqueous, ethanol and chloroform at 80°C, 40°C and room temperature respectively^{15,16}. The dried extract was stored at 4°C until use. The extract yields of the plants were 1.2, 3.0 and 2.0g from 20.0, 30.0 and 20.0g of

powered leaves in 150ml water, 300ml methanol and 250ml chloroform respectively. The aqueous extract was dissolved in 0.9% saline while the methanol extracts and chloroform extract were dissolved in 2.5% Tween 80 and subsequently in normal saline.

Wister rats of either sex and of approximately the same age, weighting about 150-175g were used for the study. They were housed in polypropylene cages are fed with standard clow diet and water *ad libitum*. The animals were exposed to alternate cycle of 12h of darkness and light. Before each test, the animals were fasted for at least 12h. The experimental protocols were subjected to the scrutinization of the Institutional Animal Ethical Committee and cleared by the same.

The animals were divided into control and test groups containing six animals each. The Control group received the vehicle (1% acacia) while the test groups received different extracts orally and were observed for mortality till 48h and the LD₅₀ were calculated¹⁷. Anti-inflammatory activity was assessed by the method described by Winter *et al.*¹⁸The rats were divided into eleven groups of six animals each. First group (negative control) received 1ml of normal saline, second group (positive control) received

10 mg/kg p.o. Diclofenac sodium, third, fourth and fifth groups were received aqueous, methanolic and chloroform extracts (100mg /kg p.o.) of *Calotropis gigantea* respectively. Groups sixth, seventh and eighth were received aqueous, methanolic and chloroform extracts (100 mg/kg, p.o) of *Tylophora indica* respectively. Groups ninth, tenth and eleventh were received aqueous, methanolic the chloroform extracts (100 mg/ kg p.o) of *Sarcostemma sacomone* respectively. After 1h, the rats were challenged with subcutaneous injection of 0.1 ml of 1% w/v solution of carrageenan into the plantar side of the left hind paw. The paw was marked with ink at the level of lateral malleolus and immersed in mercury up to the mark. The plethysmograph apparatus used for the measurement of rat paw volume was that of Singh and Ghosh¹⁹. The paw volume was measured immediately after injection (0h) and then first, second and third hour after injection of carrageenan to each group. The difference between the initial and subsequent reading gave the actual edema volume. Percent inhibition of inflammation was calculated using the formula, Percent inhibition = $100 (1 - V_t / V_c)$, where 'Vc' represents edema volume in control and 'Vt' edeam volume in group treated with various plant extracts. The

data were statistically analyzed through student's t-test.

RESULTS AND DISCUSSION

In the present study, anti-inflammatory activity of aqueous, methanol and chloroform extracts of *C. gigantea*, *T.indica* and *S.secomone* leaves were evaluated. From the results it was noted that the extracts significantly ($P < 0.05$) inhibited the inflammatory edema, though the inhibition was highest in methanolic extracts. The effect of methanolic extract was the same as that of 150mg/kg of Diclofenac sodium, among the three tested plant species *S.secomone* showed the maximum inhibition. Extracts of *C. gigantea*, *T.indica* showed a similar anti-inflammatory effect but lower than extract of *S.secomone*.

Carrageenan-induced rat paw edema has been used as an inflammation model in order to investigate the anti-inflammatory effect of drug²⁰. Carrageenan induced inflammation is a biphasic phenomenon. The first phase of edema is attributed to release of histamine and 5-hydroxytryptamine. Plateau phase is maintained by kinin like substances and second accelerating phase of swelling is attributed to prostaglandin like substances. The knowledge of these mediators involved in different phases is important

for interpreting mode of drug action^{21, 22, 23}. The result of the present study has shown that all the crude extracts of the investigated three plants exhibited very high anti-inflammatory activities. These activities may be linked with presence of polyphenolic compounds present in the extract. On preliminary phytochemical screening showed that the extract of *C. gigantea*, *T.indica* and *S.secomone* contained anti-oxidative constituents, which includes flavonoids or flavonoidal compounds. Flavonoids found in many plants have been shown to have diuretic, laxative, antispasmodic, anti-hypertensive and anti-inflammatory actions^{24, 25}.

From the results it has been concluded that the leaves of *C. gigantea* and *S.secomone* possess significant anti-inflammatory property in rats. There is increasing evidence that lysosomal enzymes play an important role in the development of acute and chronic inflammation^{26,27,28,29}. Most of the anti-inflammatory drugs exert their beneficial effects by inhibiting either release of these enzymes or by stabilizing lysosomal membrane, which is one of the major events responsible for the inflammatory process³⁰. So, we can assume that our drug extract might be acting by either inhibiting the lysosomal enzymes or stabilizing the membrane. From the above studies it is

quite apparent that the ethanolic extract possesses significant anti-inflammatory activity. The study justifies its use in inflammation as suggested in the folklore medicines. Further studies involving the purification of the chemical constituents of

the plant and the investigation on the biochemical pathways may result in the development of a potent anti-inflammatory agent with a low toxicity and higher therapeutic index.

TABLE 1. Anti-inflammatory activity of aqueous extract of *C. gigantea T.indica* and *S.secomone* plants on carrageenan induced rats hind paw edema model.

Treatment	Dose (Mg/Kg, P.O.)	Mean Paw Volume					% inhibition of edema
		0 hr	1 hr	2 hr	3 hr	4 hr	
Control	00	9.0± 0.21	9.1± 0.14	10.3± 0.05	11.4± 0.13	11.6± 0.11	-
Standard (Diclofence sodium)	150	7.6± 0.01	7.3± 0.12	6.6± 0.17	5.9± 0.11	4.2± 0.12	63.78
<i>Calotropis gigantea</i>	100	6.3± 0.04	6.8± 0.11	6.4± 0.11	6.2± 0.18	5.6 ± 0.12	51.62
<i>Tylophora indica</i>	100	7.7± 0.05	8.1± 0.13	7.4± 0.16	6.2± 0.19	5.2± 0.13	55.27
<i>Sercostemma secomone</i>	100	7.5± 0.12	8.3± 0.13	7.2± 0.14	6.0± 0.04	5.5± 0.10	52.63

TABLE 2. Anti-inflammatory activity of ethanol extracts of *C. gigantea T.indica* and *S.secomone* plants on carrageenan induced rats hind paw edema model.

Treatment	Dose (Mg/Kg, P.O.)	Mean Paw Volume					% inhibition of edema
		0 hr	1 hr	2 hr	3 hr	4 hr	
Control	00	9.0± 0.21	9.1± 0.14	10.3± 0.05	11.4± 0.13	11.6± 0.11	-
Standard (Diclofence sodium)	150	7.6± 0.03	7.3± 0.12	6.6± 0.17	5.9± 0.11	4.2± 0.12	63.81
<i>Calotropis gigantea</i>	100	7.2± 0.12	8.1± 0.11	6.6± 0.12	6.4± 0.14	4.8 ± 0.23	58.73
<i>Tylophora indica</i>	100	8.3± 0.16	9.8± 0.23	7.5± 0.18	7.3± 0.11	6.6± 0.12	43.28
<i>Sarcostemma secomone</i>	100	7.5± 0.11	8.4± 0.15	7.3± 0.21	6.1± 0.24	4.7± 0.23	40.62

TABLE 3. Anti-inflammatory activity of chloroform extract of *C. gignatea* *T.indica* and *S.secomone* plants on carrageenan induced rats hind paw edema model.

Treatment	Dose (Mg/Kg, P.O.)	Mean Paw Volume					% inhibition of edema
		0 hr	1 hr	2 hr	3 hr	4 hr	
Control	00	9.0± 0.21	9.1± 0.14	10.3± 0.05	11.4± 0.13	11.6± 0.11	-
Standard (Diclofence sodium)	150	7.6± 0.03	7.3± 0.13	6.6± 0.17	5.9± 0.11	4.2± 0.12	63.85
<i>Calotropis gigantea</i>	100	7.1± 0.11	6.9± 0.14	6.6± 0.11	6.0± 0.12	5.4 ± 0.16	53.59
<i>Tylophora indica</i>	100	7.8± 0.17	6.7± 0.14	6.3± 0.06	5.6± 0.14	5.3± 0.12	54.42
<i>Sarcostemma secomone</i>	100	7.5± 0.05	6.4± 0.08	6.2± 0.11	5.3± 0.05	4.8± 0.11	58.61

REFERENCES

1. Katzung, B.G. Basic and Clinical Pharmacology. Lang Medical Books, McGraw-Hill, New York, **2001**; 536-539.
2. Satoskar, S.D, Bhandarkar, S.S. Pharmacology and pharmacotherapeutics. Popular Prakashan Pvt. Ltd, Mumbai, **2005**; 153-160.
3. Smith, M.C. and Reynard, M. A. Essential of Pharmacology. Saunders Company, West Bengal, **1995**; 147-149.
4. Pramod K.Tyagi, Vishnu K. Rai, Ashish K.Pahria, S. Sambath Kumar, Yogendra Singh, Manoj Sharma. and Manoj Goyal. Preliminary phytochemical screening and evaluation of anti-inflammatory activity of ethanolic extract of leaves of *Indigofera tinctoria* Linn. *Journal of Current Pharmaceutical Research*, **2010**; 3(1): 47-50.
5. Sosa S, Balick M.J. Arvigo R, Esposito R.G. Pizza C, Altiniez G. A screening of the topical anti-inflammatory activity of some Central American plants. *J. Ethanopharmacol*, **2002**; 8: 211-215.
6. Ravikumar, A, Subburathinam, K.M. and Prabakar, G. Studies on anti-inflammatory property of *Peltophorum pterocarpum*, *Colvillea rocemosa* and *Bauhinia*

- purpurea* leaves in experimental animal models. *Adv. Pharmacol. Toxicol*, **2006**; 7(3): 13-16.
7. Patil, V.V. Pimprikar, R.B. and Patil, V.R. Pharmacognostical studies and evaluation of anti-inflammatory activity of *Ficus bengalensis* linn. *Pharmacognosy*, **2009**; 1 (1): 49-53.
 8. Alam, K., D. Pathak and S. H. Ansari. Evaluation of anti-inflammatory activity of *Adhatoda zeylanica* (medic.) leaves extract. *International Journal of Pharma and Bio Sciences*, **2011**; 2 (1): 157-162.
 9. Bagul, M.S, Shrinivas, H, Kanaki, M.S. and Rajani, M. Anti-inflammatory activity of two ayurvedic formulation containing guggul. *Ind. J. Pharmacol*, **2005**; 37: 299-400.
 10. Bhattacharya, P, Lakshmi, S.M, Ashok Kumar, C.K. and Mandal, S.C . Conference proceeding on Promotion and Development of Botanicals with International Coordination School of National Product, Jadavpur, Kolkota, **2005**; 383-385.
 11. Amberkar Mohanbabu V. R, Tara Shanbhag Meena Kumari, K, K. L. Bairy and Smita Shenoy. Evaluation of anti-inflammatory and analgesic activities of alcoholic extract of *Kaempferia galanga* in rats. *Indian J. Physiol. Pharmacol*, **2011**; 55 (1): 13-24.
 12. Mahesh, S. Paschapur, M. B. Patil, Ravi Kumar, and Sachin R. Patil. Evaluation of anti-inflammatory activity of ethanolic extract of *Borassus flabellifer* L. male flowers (inflorescences) in experimental animals. *Journal of Medicinal Plants Research*, **2009**; 3(2): 49-54.
 13. Sudipta Das, Pallab, K. Haldar, Goutam Pramanik. and R.B. Suresh. Evaluation of anti-inflammatory activity of *Clerodendron infortunatum* Linn. extract in rats. *Global Journal of Pharmacology*, **2010**; 4 (1): 48-50.
 14. Rajamanickam, E, Gurudeeban, S, Ramanathan, T. and Satyavani, K. Evaluation of anti-inflammatory activity of *Citrullus colocynthis*. *International Journal of Current Research*, **2010**; 2: 67-69.
 15. Kokate, C.K. Practical Pharmacology, 3rd Edn, Vallabh Prakashan, New Delhi, **1994**; pp: 153-158.

16. Owoyele, B. V. Olaley, e S. B., Oke, J.M. and Elegbe, R. A. . Anti-Inflammatory and analgesic activities of leaf extracts of *Landolphia oweriensis*. *Afr. J. Res*, **2001**; 4: 132-133.
17. Ghosh, M.N. Fundamental of Experimental Pharmacology, 2nd Edn, Scientific book agency, Kolkata, **1994**; 153-158.
18. Winter C.A. Risley E.A, and Silber R.H. Anti-inflammatory activity of indomethacin and plasma corticosterone in rats. *J. Pharmacol Exp. Ther*, **1968**; 113: 693-698.
19. Singh, H. and Ghosh, M.N. Modified plethysmometer for measuring foot volume of anaesthetized rats. *J. Pharm Pharmacia*, **1968**; 20: 316-317.
20. El-Shenawy S.M. Abdel -Salam O.M. Baluomy A.R. El-Batran S. and Arbid M.S. Studies on the inflammatory and anti-nociceptive effects of melatonin in the rat. *Pharmacol. Res.*, **2002**; 46: 235-243.
21. Muthumani, P, R. Meera, P. Devi1, Shiny george, S.A. Mohamed Shiek, Arabath, K. Jeyasundari and R. Babmanaban. . Phytochemical investigation, diuretic and anti-Inflammatory activity of root and stem extracts of *Boerhaavia erecta* Linn in experimental animals. *International Journal of Applied Biology and Pharmaceutical Technology*, **2010**; 1285-1292.
22. Goodman and Gilman. The Pharmacological basis of therapeutics, McGraw Hill, New York, 10th ed, **2001**; 687.
23. Moulisha Biswas, Kaushik Biswas, Tarun K. Karan, Sanjib Bhattacharya, Ashoke, K. Ghosh. and Pallab K. Haldar. Evaluation of analgesic and anti-Inflammatory activities of *Terminalia arjuna* leaf. *Journal of Phytology and Phytopharmacology*, **2011**; 3(1): 33-38.
24. Nayak, S. Sanjay Jain., Mohammad Ayub. and Singhai A.K. Anti-inflammatory activity of the bark of *Elaeodendron glaucum*, Peers. *Adv. Pharmacol Toxicol*, **2006**, 7(1): 51-53.
25. Mule, S. N., Ghadge, R. V., Chopade, A. R., Bagul, B.A., Patil, S. B. and Naikwade, N. S. Evaluation of antinociceptive and anti-inflammatory activity of leaves of gynandropsis

- pentaphylla. *Journal of Herbal Medicine and Toxicology*, **2008**; 2 (1) 41-44.
26. Anderson, A.J, Bocklehurst, W.E and Wills, A.L. Evidence for the role of lysosomes in the formation of prostaglandins during carraginin induced inflammation in rat. *Pharmacol. Res. Comm*, **1971**; 3: 13-17.
27. Shen, T.Y. . In: Robinowitz, Myerson RM (eds). *Topics in Medicinal Chemistry*, Vol. 1, Wiley InterScience, New York. **1967**; pp: 29-38.
28. Weissmann, G. The role of lysosome in inflammation and disease. *A. Rev. Med*, **1967**; 18: 97-101.
29. Jannoff, A. and Zweifach, B.W. Production of inflammatory changes in the micro-circulation by cationic proteins extracted from lysosomes. *J. Exp. Med*, **1964**; 120: 747-752.
30. Nair, R.B, Ravishankar, B, Vijayan, N.P, Sasikala, C.K and Saraswathy, V.N. Anti-inflammatory effect of *Strbilanthus heyneanus* leaves- A biochemical study. *J. Res. Ay. Sid*, **1998**; 9: 46-50.